RESEARCH ARTICLE

A Novel Method for the Selective Identification of Metformin in Bulk and Pharmaceutical Formulations Using Sodium Nitroprusside as a Chromogenic Reagent by Spectrophotometry

SHIVAJI R. LABHADE^{1*}, KAILAS R. LABHADE² and VISHWAS B. GAIKWAD³

¹Department of Chemistry, K. T. H. M. College, Gangapur Road, Shivajinagar, Nashik- 422002, India

²Department of Chemistry, S.V.K.T. Arts, Science and Commerce College, Deolali Camp, Nashik- 422401, India

³Director, BCUD, University of Pune, Ganeshkhind, Pune-411007, Maharashtra State, India *srlabhade3571@rediffmail.com*

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Abstract: A novel method to identify the metformin with a good selectivity has been established by using sodium nitroprusside as a chromogenic reagent. The experiment indicates that in basic solution sodium nitroprusside can react with the oxidized product (obtained by the action of sodium hypochlorite) of metformin to form a green colored 1, 3 dinitrosyl-*N*'-(iminomethyl)-*N*, *N*-dimethyl formamidine with maximum absorption at 680 nm. The test when compared with blank is sensitive for spectrophotometric detection of metformin to sodium nitroprusside is 1:2. The reaction stoichiometric ratio of the oxidized product is discussed in present investigation. The proposed method has been successfully applied for identification of metformin in aqueous solution containing glibenclamide, glimepiride, glipizide and gliclazide. Because of the dissociative nature of the green colored product is not found to be linear. Hence it can concluded from this analysis that sodium nitroprusside is not suitable chromogenic reagent for spectrophotometric determination of metformin in aqueous solution nitroprusside is not suitable chromogenic reagent for spectrophotometric determination of metformin in aqueous solution nitroprusside is not suitable chromogenic reagent for spectrophotometric determination of metformin in aqueous solution nitroprusside is not suitable chromogenic reagent for spectrophotometric determination of metformin in aqueous solution.

Keywords: Metformin, Metformin Identification, Sodium Nitroprusside, Sodium Hypochlorite

Introduction

Metformin is an oral antihyperglycemic agent used to lower blood glucose in patients suffering from non-insulin dependent diabetics mellitis¹. Chemically metformin is N, N-dimethylimidodicarbonimidic diamide hydrochloride² with molecular weight 165.62 g/mol.

Metformin hydrochloride

Spectrophotometry is the most common and convenient analytical tool due to its inherent simplicity and low costing. Thus is used in most of the quality control and clinical laboratories. Literature survey indicates that several spectrophotometric methods are used for determination of metformin. These includes direct determination with multivariate calibration technique³, reaction of metformin with cupric ion and cyclohexylamine in basic medium⁴, formation of oxidative coupling of the reaction product of metformin with 1-naphthol⁵, oxidation of metformin yellow-colored chromophore⁶, formation of charge transfer complex of metformin with iodine in acetonitrile medium⁷ as well as determination with the first-derivative method⁸⁻⁹. There is no any visible spectrophotometric method yet reported for quantitation of metformin using sodium nitroprusside. The aim of this study was to develop a new spectrophotometric method for quantitative determination of metformin as a raw material and in some pharmaceutical preparations with no interference of other constituents in their formulations.

Sodium pentacyanonitrosylferrate(II) or sodium nitroprusside (SNP), Na₂[Fe(CN)₅NO] is a popular derivatization agent of analytically important for the detection and determination of wide variety of nucleophilic moieties such as primary or secondary aliphatic amines, aldoximes, ketones, nitrites, phenols, pyrroles, quinones, thiols, thioureas and uracils¹⁰⁻¹¹. Many spectrophotometric methods adapted SNP as an analytical reagent in their applications, including the assays of Rossoxacin¹², Paracetamol¹³, Streptomycin¹⁴, Meloxicam¹⁵, Cefradine¹⁶, Diosmin¹⁷, Atenolol¹⁸, Methanol¹⁹, Piroxicam²⁰, Sumatriptan succinate²¹, Ramipril²² and Thiosulphate²³. Furthermore, the techniques like high performance liquid chromatography²⁴ and flow injection analysis²⁵ have also adapted SNP as an analytical reagent in their applications. In addition, the coupling reaction of SNP with the diazotized *p*-nitroaniline in basic solution also been used for spectrophotometric determination of sodium nitroprusside²⁶. For detection of impurities²⁷ with TLC technique, the mixture of SNP, potassium ferrocyanide and sodium hydroxide has been employed for identification of metformin. It has an added advantage that the test does not involved oxidation of metformin by sodium hypochlorite solution. Spectrophotometric determinations of metformin⁵⁻⁶ and paracetamol²⁸ have been utilized sodium hypochlorite used as oxidizing reagent for oxidation of both the analytes. It has been reported that using sodium hypochlorite solution, metformin is oxidized to its β -diketone⁶ product which subsequently reacts with SNP in basic solution. Due to nucleophilic reactivity of oxygen atoms of the β diketone of metformin with electrophilic nitrosyl group in two SNP molecules, a green colored 1, 3 dinitrosyl-N'-(iminomethyl)-N, N-dimethyl formamidine complex is formed. Because of dissociative nature of this reaction product we were unable to develop a method for quantitative determination of metformin. But using SNP as chromogenic reagent same experiment gives a novel method for selective identification of metformin in aqueous solution containing glibenclamide, glimepiride, glipizide and gliclazide is discussed in this paper.

Experimental

Shimadzu UV-1800 double beam UV-Vis spectrophotometer with software UVProbe 2.33 using 10 mm quartz silica cells was used for all spectral measurements.

Reagents

All the chemicals used were of analytical grade and so used without further purification. Distilled water was used for diluting the standard, reagents and samples. All the prepared reagents were stored in the amber colored bottles and protected from direct sunlight.

Sodium hypochlorite (NaOCl) solution

A 0.2% solution was prepared by diluting 25.0 mL of 4% solution of commercially available sodium hypochlorite to 500 mL with distilled water and stored in amber colored bottle with air tight stopper.

Sodium nitroprusside (SNP) solution

A 2.0 x 10^{-2} mol L⁻¹ solution was prepared by dissolving 2.980 g of Na₂[Fe(CN)₅NO].2H₂0 in 500 mL of distilled water.

Sodium hydroxide (NaOH) solution

A 1.0 mol L^{-1} solution was prepared by dissolving 20.0 g of NaOH in 500 mL of distilled water.

Standard metformin hydrochloride (MHCl) solution

A 1.0×10^{-3} mol L⁻¹ of MHCl solution was prepared by dissolving 82.81 mg of pharmaceutical grade MHCl (C₄H₁₁N₅HCl, 165.62 g mol⁻¹) in 500 mL of distilled water.

Dosage forms

Tab. Metfor (500 mg MHCl), Tab. Glycomet (500 mg MHCl), Tab. Glycinorm (80 mg gliclazide), Tab. Daonil (5 mg glibenclamide), Tab. Glimestar (1 mg glimepiride) and Tab. Glyzip (5 mg glipizide)

General Analytical Procedure

Caution

Potentially hazardous volatile gasses were possibly evolved during the experimentation. Preparation of solutions of metformin with sodium hypochlorite and sodium nitroprusside was done in a fuming hood. Absorbance measurement was carried out in a well ventilated area.

Aliquots of 5.0 mL of aqueous solution of MHCl containing 16.56-82.80 μ g of MHCl were transferred to 50 mL beakers containing 2.0 mL of 1.0 mol L⁻¹ NaOH solution. Then 1.0 mL of 0.2% NaOCl solution and 2.0 mL of 5.0 x 10⁻³ mol L⁻¹ of SNP solution were added. In absence of direct sunlight the reaction mixtures were made to react at room temperature for 2 minutes. Later, according to the sequential addition of SNP reagent, the absorbance of the green colored complex was instantaneously measured at 680 nm against reagent blank. The increase in absorbance of the test solution with respect to blank indicates presence of metformin in the solution. For test solution having concentration of MHCl greater than and equal to 49.68 μ g, the green color thus produced by this reaction was noticeable without instrument. The green color obtained only in strongly basic solution and found to be completely vanished when solution was made neural or acidic.

Results and Discussion

The aim of this work is to establish a simple, sensitive, reliable, selective and cheap spectrophotometric method for the identification of metformin drug in pure form and in its pharmaceutical formulations.

Reaction mechanism and reaction stoichiometry

The developed method is based on reaction of metformin with the sodium hypochlorite and SNP reagents in basic solution. Addition of sodium hypochlorite to the alkaline solution of metformin hydrochloride yields β -diketone⁶, an oxidized product of the metformin. Sodium

nitroprusside can react with many nucleophilic agents in basic solution and form the colored derivative due to electrophilic reactivity of its nitrosyl group¹⁰⁻¹¹. Similarly, in basic medium the oxygen atoms of β -diketone molecule of metformin have nucleophilicity and trend to attack on the electron deficiency center in nitrosyl groups of two SNP molecules to give 1, 3 dinitrosyl-*N*⁻(iminomethyl)-*N*,*N*-dimethyl formamidine compound. It was proved by varying the concentration of metformin and SNP, the stoichiometric ratio of reaction between β -diketone of the metformin and SNP is 1:2. It is presumed that both the oxygen of the β -diketone of metformin reacts with nitrosyl groups. Hence, it seems reasonable that the reaction mechanism is as follows (Scheme 1):



Scheme 1. Reaction mechanism

The reaction reported for determination of paracetamol¹³ illustrate that, paracetamol molecule linked with nitrosyl group through phenolic oxygen gives green colored compound with absorption maxima at 700 nm. Similarly, β -diketone of metformin reacts with two nitrosyl groups of two SNP molecules through the two oxygen atoms also gives the green color to the product because of the oxygen-nitrosyl group linkages. The order of addition of reagents affects the results of identification of metformin. The order used in the proposed method was aqueous solutions of metformin, sodium hypochlorite and SNP. Without pre-reaction by sodium hypochlorite solution metformin does not give the green color with sodium nitroprusside.

Selection of wavelength

This 1,3 dinitrosyl-N'-(iminomethyl)-N, N-dimethyl formamidine complex of β -diketone of metformin exhibits an absorption spectrum as shown in Figure 1 with maximum absorption at 680 nm. The existence of peak at 680 nm for the colored product is attributed to the

bathochromic shift because of increase in conjugation in the β -diketone molecule due to substitution of two nitrosyl groups. The β -diketone of metformin showed inertness towards the potassium ferrocyanide consequently cyanide ligand of the nitroprusside anion does not react with β -diketone anion.



Figure 1. Absorption spectra (obtained against distilled water) at 10 mL dilution of 2.0 mL of 0.5×10^{-3} mol L⁻¹ of pure MHCl (1) of 1.0 mL of 0.2 % pure NaOCl solution (2) of 2.0 mL of 1.0 mol L⁻¹ of NaOH (3) of 2.0 mL of 2.0 $\times 10^{-2}$ mol L⁻¹ of pure SNP (4) and of 0.0, 1.0, 2.0, 3.0 and 4.0 mL of 0.5 $\times 10^{-3}$ mol L⁻¹ of MHCl (5-9) added to 2.0 mL of 1.0 mol L⁻¹, 1.0 mL of 1% NaOCl of NaOH and 2.0 mL of 2.0 $\times 10^{-2}$ mol L⁻¹ of SNP.

Aqueous solution of MHCl, NaOCl, NaOH, and SNP does not show absorbance at the wavelength at which 1, 3 dinitrosyl-*N*'-(iminomethyl)-*N*, *N*-dimethyl formamidine complex of β -diketone of metformin exhibits maximum absorption. Consequently, the spectral region 650 to 750 nm was found to be appropriate for detection of metformin through the absorbance measurement. The wavelength of maximum absorption (λ -max) of the green colored product and its color intensity were found fluctuated with the concentrations of metformin and SNP. For lower concentration of metformin as well as sodium nitroprusside, the absorption maximum was found shifted to longer wavelength.

Color stability of the product

Aqueous solution of metformin after reaction with NaOCl, SNP in NaOH produces a green colored derivative product of the β -diketone of metformin. The absorbance of the formed product was measured at different time interval. It was found that color intensity of the green colored product was found to be gradually increasing to maximum within the time span of 2 to 3 minutes and thereafter decreasing to minimum as shown in Figure 2.

Under the recommended conditions, there is a linear relationship between the absorbance and the concentration of metformin. For different solutions having microgram concentration of metformin, absorbance of the green colored product is not linear with the concentration of metformin because of the uneven time span in addition of SNP reagent and measurement of the absorbance. Because of dissociative nature of the reaction product, the absorbance of the same solution when measured at different time interval produces different

absorbance readings. Consequently, the time span laps in between the absorbance measurement of a series of solutions produces non-linear relation between absorbance and concentration of metformin. Therefore, the procedure was not found suitable for quantitative determination of metformin in aqueous medium.



Figure 2. Absorption spectra (obtained against reagent blank) at 10 mL dilution of 3.0 mL of 0.25×10^{-3} mol L⁻¹ of pure MHCl added to 2.0 mL of 1.0 mol L⁻¹ of NaOH, 1.0 mL of 1% NaOCl, and 3.0 mL of 5.0×10^{-3} mol L⁻¹ of SNP. After addition of SNP reagent, the curves 1 to 11 were obtained at 1, 3, 5, 7, 9, 11, 13, 15, 17, 19 and 21 minutes respectively.

Sensitivity of detection

The procedure is suitable for detection of metformin hydrochloride in aqueous solution up to concentration of 16.56 μ g at 10 mL dilution. Figure 3 illustrates that reagent blank solution is transparent at the wavelength of analysis. The test solution containing metformin showed increase in absorbance. For low concentration of metformin, the absorbance was found negligible and detection of metformin in the solution was found only by spectrophotometer. But solutions having concentration of metformin greater than and equal to 49.68 μ g showed distinct visible green color and detection of metformin was feasible without spectrophotometer.



Figure 3. Absorption spectra (obtained against distilled water) at 10 mL dilution of 0.0, 16.56, 33.12, 49.68 and 66.24 μ g of MHCl (1-5) added to 2.0 mL of 1.0 mol L⁻¹ of NaOH, 1.0 mL of 0.2% NaOCl, and 2.0 mL of 5.0 x 10⁻³ mol L⁻¹ of SNP.

Interference study in detection of metformin

The effect of ingredients commonly found in metformin formulations was studied by the identification of metformin with the addition of ingredients. The commercially available dosage forms of metformin²⁹ contain pure metformin and includes Metfor and Glycomet tablets each with 500 mg MHCl. In addition to this dosage forms of the metformin is also available in combination with glibenclamide (Tab. Diabetrol: 500 mg MHCl + 5 mg glibenclamide), gliclazide (Tab. Reclimate: 500 mg MHCl + 80 mg gliclazide), glipizide (Tab. Actizide-M: 500 mg MHCl + 5 mg glipizide), glimepiride (Tab. Gemer 1: 500 mg MHCl + 1 mg glimepiride), etc. In order to study the interfering role of glibenclamide, gliclazide, glipizide and glimepiride in metformin detection, the alkaline solutions of the tablet Daonil (5 mg glibenclamide), Glycinorm (80 mg gliclazide), Glyzip (5 mg glipizide), and Glimestar (1 mg glimepiride) were tested by above reaction. The solution of each single tablet was prepared separately in 25.0 mL of 1.0 mol L⁻¹ of NaOH and after filtration through Whatman filter paper, 2.0 mL of solution of each tablet was tested by above recommended procedure. It was observed that none of the above mentioned in alkaline solution could produce green color made to react with sodium hypochlorite and sodium nitroprusside. The absorption spectra Figure 4, illustrates that reaction products of glibenclamide, glimepiride, glipizide, gliclazide obtained with sodium hypochlorite and sodium nitroprusside in basic medium are transparent at the wavelength of analysis. Furthermore, the interference of four tablets was also tested by same procedure with their dissolution in 25.0 mL of 95% ethanol. It was noted that all tablets showed negative test with sodium hypochlorite and sodium nitroprusside.



Figure 4. Absorption spectra (obtained against reagent blank) at 10 mL dilution of aqueous solution of glibenclamide, glimepiride, glipizide, gliclazide (1-4), 82.81 and 165.62 μ g of MHCl (5-6), added to 2.0 mL of 1.0 mol L⁻¹ of NaOH, 1.0 mL of 1% NaOCl, and 2.0 mL of 5.0 x 10⁻³ mol L⁻¹ of SNP.

Detection of metformin in synthetic pharmaceutical formulations

Detection of metformin in presence of gliclazide, glibenclamide, glimepiride or glipizide was carried out by forming their mixture with metformin. A single tablet of gliclazide (80 mg), glibenclamide (5 mg), glimepiride (1 mg) or glipizide (5 mg) was powered and dissolved in 25 mL of 95% ethanol. The solution was filtered through Whatman filter paper 42. Later 5.0 mL filtrate of gliclazide, glibenclamide, glimepiride or glipizide tablet was added to pure metformin hydrochloride solution and diluted to 100 mL with distilled water so

that final solution was contain 20.0 μ g mL⁻¹ of metformin with other constituent. This solution was then subjected for detection of metformin by obtaining the absorption spectra against distilled water.

The result of detection of metformin in presence of gliclazide is shown in Figure 5, which illustrate that, though the alcoholic solution of gliclazide does not generate green color on reaction with sodium nitroprusside, its presence with metformin still decreases formation of green color complex. The shift in absorption maxima to the shorter wavelength and decrease in color intensity of the product indicates that gliclazide generates colorless product on reaction with these reagents. The result of detection of metformin in presence of glibenclamide, glimepiride and glipizide are reported in Figure 6, Figure 7 and Figure 8 respectively. These results interpret that, alcoholic solutions of glibenclamide, glimepiride and glipizide in presence of metformin does not affects the green color intensity of the product.



Figure 5-8. Absorption spectra of 0.0, 20.0, 40.0, 60.0, 80.0 and 100.0 µg of MHCl (1-6) spiked with gliclazide (Figure 5), glibenclamide (Figure 6), glimepiride (Figure 7) and glipizide (Figure 8) respectively.

In addition the detection of metformin was also carried out with the mixture of gliclazide, glibenclamide, glimepiride and glipizide. An aliquots of 1.0 mL alcoholic filtrate of each tablet (prepared as above) were added to pure MHCl solution and diluted to 80 mL with distilled water so that final solution was containing 20.0 μ g ml⁻¹ of metformin. Different aliquots of this diluted solution were subjected for detection of metformin. The result for detection of metformin in presence of all these four constituents is shown in Figure 9,

which illustrates that the mixture of gliclazide, glibenclamide, glimepiride and glipizide does not affect the intensity green color and the absorption maxima of the product. This could be probably due to low concentration of gliclazide in this mixture.



Figure 9. Absorption spectra (obtained against distilled water) at 10 mL dilution of 0.0, 20.00, 40.00, 60.00, 80.00 and 100.0 μ g of MHCl (1-6) spiked with the mixture of gliclazide, glibenclamide, glimepiride and glipizide.

Conclusion

The proposed spectrophotometric procedure is simple, cost-effective, time saving and permits direct identification of metformin in the aqueous solution containing glibenclamide, glimepiride, glipizide and gliclazide. Sodium nitroprusside is able to reacts in basic medium at room temperatures with the oxidized product of the metformin to produce green colored 1, 3 dinitrosyl-N'-(iminomethyl)-N, N-dimethyl formamidine complex. Consequently, the suggested method is selective and free from interferences of the common excipients and additives present in pharmaceutical dosage forms. Moreover, it exhibits the advantage of utilization of a single step reaction and sensitive for spectrophotometric detection of metformin upto 16.56 µg. The detection of metformin is also feasible without spectrophotometer when concentration of metformin greater than and equal to 49.68 µg in 10 mL solution. The method should be useful for routine analytical and quality control identification of the metformin. The green colored 1, 3 dinitrosyl-N'-(iminomethyl)-N, Ndimethyl formamidine complex of the oxidized product of metformin was obtained only in strong basic solution and has dissociative nature in aqueous medium. Consequently it can also be concluded that the procedure is not suitable for quantitative determination of metformin in aqueous medium.

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References

1. British Pharmacopoeia. Vol II, Cambridge University Press, Cambridge, 1988.

- 2. Bailey C J and Path M R C and Robert C Turner *New Engl J Med.*, 1996, **334(9)**, 574-579.
- 3. Arayne M S, Sultana N, Zuberi M H and Siddiqui F A, *Indian J Pharm Sci.*, 2009, **71**, 331-335; DOI:10.4103/0250-474X.56022
- 4. Hassan S S M, Mahmoud W H, Elmosallamy M A and Othman A H, *Anal Chim Acta* 1999, **378**, 299-311.
- 5. Ahmad N R and Omar F K, Iraqi National J Chem., 2012, 46, 161-170.
- 6. Ahmad N R, Iraqi J Pharm., 2012, **12**, 75-85.
- 7. Ashour S and Kabbani R, *Anal Lett.*, 2003, **36(2)**, 361-370; DOI:10.1081/AL-120017696
- Onal Armagan, *European J Med Chem.*, 2009, 44(12), 4998-5005; DOI:10.1016/j.ejmech.2009.09.003
- 9. Thomas A B and Patil S D, *Der Pharma Chemica*, 2011, **3**(3), 271-276.
- Rucki R, In: K. Florey (Ed.), Analytical Profile of Drug Substances, Vol. 6, Academic Press, New York, 1977, pp. 488-513.
- 11. Leeuwenkamp O R, Van Benneekom W P and Bult A, In: Florey K, (Ed.), Analytical Profile of Drug Substances, Vol. 15, Academic Press, New York, 1986, pp. 782-789.
- 12. Askal H F, Refaat I H, Darwish I A and Marzouq M A, *Spectrochimica Acta Part-A: Mole Biomole Spectr.*, 2008, **69**, 1287-1291; DOI:10.1016/j.saa.2007.07.031
- 13. Yanyan Zhan, Yan Zhang, Quanmin Li and Xinzhen Du, J Anal Chem., 2011, 66, 215-220.
- 14. Quan-min Li and Lin-xiao Gao, Anal Lett., 2008, **41(14)**, 2595-2607; DOI:10.1080/00032710802363354
- 15. Gurupadayya B M, Trinath M N and Shilpa K, *Indian J Chem Technol.*, 2013, **20**, 111-115.
- Hua Zhang, Ling Li Wu, Quan Min Li and Xin Zhen Du, *Chinese Chem Lett.*, 2008, 19(12), 1470-1474; DOI:10.1016/j.cclet.2008.09.014
- Zenovia Moldovan and Hassan Y Aboul-Enein, *Instru Sci Technol.*, 2011, **39(2)**, 135-148; DOI:10.1080/10739149.2010.545851
- 18. Bhashir N, Shah S W H, Bangesh M and Riazullah, J Sci Indu Res., 2011, 70, 51-54
- Yan-Yan Zhan, Yan Zhang, Quan-Min Li and Xin-Zhen Du, J Chine Chem Soc., 2010, 57(2), 230-235; DOI:10.1002/jccs.201000035
- 20. Rele R V, Sawant S A and Warkar C B, Int J Chem Tech Res., 2010, 2(4), 2173-2176.
- 21. Kalyanaramu B and Raghubabu K, Der Pharma Chemica, 2011, 3(1), 223-228.
- 22. Kalyanaramu B, Raghubabu K and Vamsikumar Y, *J Chem Pharm Res.*, 2011, **3**(2), 863-869.
- 23. Sharma R, Rathore S S, Sharma P and Sharma A, *J Chem Pharmace Res.*, 2009, **1**(1), 321-328.
- 24. El-Walily A F M, Abdine H H, Razak O A and Zamel S, *J Pharm Biomed Anal.*, 2000, **22(6)**, 887-897; DOI:10.1016/S0731-7085(99)00290-3
- 25. Shah J, Rasul J M and Rehman F, J Chil Chem Soc., 2008, 53, 1605-1608.
- 26. Kasim Y.R, Batool B I and Hayfa M.J, Raf J Sci., 2011, 22(3), 119-128.
- 27. European Pharmacopoeia 5.0, Monographs M, pp. 2003-2004 and European Pharmacopoeia 7.0, Monographs M, pp.2458-2459.
- Vilchez J L, Blanc R, Avidad R and Navalon A, J Pharma Biomed Anal., 1995, 13(9), 1119-1125; DOI:10.1016/0731-7085(95)01537-U
- 29. Drug Today, India, Vol. II, Oct.-Dec.2011, pp.1079-1086.