

## Synthesis and DNA Binding of *cis*-Dichlorobis-benzimidazolebis-(dimethylsulfoxide)ruthenium(II)-hydrate Complexes

JITENDRA DEKA<sup>1</sup>, KIBRIYA SIDDIQUE<sup>2</sup>, PANKAJ HAZARIKA<sup>1</sup>,  
OKHIL KUMAR MEDHI<sup>1</sup>, SANJIB KARMAKAR<sup>2</sup> and CHITRANI MEDHI<sup>1\*</sup>

<sup>1</sup>Department of Chemistry, Gauhati University, <sup>2</sup>Department of Instrumentation & USIC-  
Gauhati University, Guwahati-781014, Assam, India  
*chitranimedhi@gmail.com*

Received 16 September 2015 / Accepted 5 October 2015

**Abstract:** The ruthenium complex with substituted imidazole ligand of formula [Ru(BIm)<sub>2</sub>(DMSO)<sub>2</sub>Cl<sub>2</sub>].H<sub>2</sub>O (BIm=Benzimidazole) was prepared. The compound has been further characterized from spectroscopic data and elemental(C, H and N) analysis. The molecular structure of this compound was determined by single crystal X-ray diffraction study. Binding of this complex with plasmid-DNA was studied by UV-Visible and Emission spectral studies. Further electrochemical studies were performed to understand the DNA binding ability of this complex. The evidences of DNA binding were found from the change of the Intensity of fluorescence band and UV-band. Binding constant ( $K_b$ ) calculated from UV-Spectroscopic titration is found to be  $9.09 \times 10^5 \text{M}^{-1}$ .

**Keywords:** Benzimidazole, DMSO, X-ray diffraction, Electrophoresis, Plasmid-DNA.

### Introduction

The imidazole-metal complexes are important in medicine<sup>1</sup> and coordination chemistry<sup>2</sup>. The imidazole and DMSO ligands have been extensively used in many metal complexes<sup>3</sup>. We have synthesized a few benzimidazole ruthenium complexes which are found to be potent anticancer agent<sup>4</sup>. Ligand's effect on anticancer property<sup>5</sup> is important and it is essential to design superior complexes in terms of effectiveness compared to that which are already available agent such as *cis*-platin. The imidazole-metal complexes are important<sup>6-9</sup> in medicine and coordination chemistry. The spectroscopy of these complexes in the absence and presence of plasmid DNA was examined and compared to *cis*-platin. Furthermore, benzimidazole complexes have been evaluated for antifungal activity and as antitumour agents<sup>10,11</sup>. Hence, the imidazole and DMSO<sup>12</sup> ligands have been extensively used many metal complexes<sup>13</sup>.

Benzimidazole cyclometalated complexes were found<sup>14</sup> to have good anticancer activity against HT29, T47D, A2780 and A2780cisR cancer cell lines<sup>15,16</sup>. Representative complexes show high apoptosis, good accumulation and S-phase<sup>17</sup> cell arrest and strongly bind to HSA at sites I and II and also weakly bind to DNA at the minor groove<sup>18</sup>. Bis-benzimidazole bridged supramolecular coordination complex has potential to act as potent anticancer agents, particularly in cell lines which are resistant to Pt-based molecules<sup>19,20</sup>. Further studies are under progress to investigate the biological mechanism of these derivatives<sup>21</sup>. We have synthesized few benzimidazole ruthenium complexes which are found to be potent anticancer agent<sup>4,22</sup>. A single crystal X-ray diffraction study of this complex is reported.

## Experimental

Analytical grade  $\text{RuCl}_3 \cdot 3\text{H}_2\text{O}$ , Plasmid-DNA (pBR 322 DNA), Tris buffer were purchased from Sigma and Aldrich chemical companies, USA.

### Characterization procedure

The IR Spectra of the compound was recorded as KBr pellets on a Perkin-Elmer FT-IR spectrophotometer. The UV-visible spectra were taken in a Shimadzu UV-2401 PC Spectrophotometer.

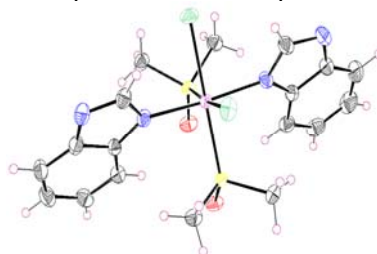
### Preparation

#### *Synthesis of cis-dichlorobis-(benzimidazole)bis-(dimethylsulfoxide)ruthenium(II) hydrate, $[\text{Ru}(\text{Blm})_2\text{D}_2\text{Cl}_2] \cdot \text{H}_2\text{O}$*

0.25 g  $\text{RuCl}_3 \cdot 3\text{H}_2\text{O}$  was refluxed in 30 mL ethanol and 5 mL, 2M HCl for 2 h, and cooled the solution. 0.1g of benzimidazole was dissolved in 6M HCl and mixed ruthenium chloride solution. The mixture was stirred for 5 min and cooled in ice for 2 h and then at room temperature for 24 h. Yellow crystals formed, Yield = 70%. Isolated crystals were separated with 1mL DMSO in ethanol for 30 min. Light yellow crystals formed. A suitable crystal was used for XRD analysis. Molecular formula  $\text{C}_{18}\text{H}_{24}\text{Cl}_2\text{N}_4\text{O}_3\text{RuS}_2$  Mol.wt = 580 g/mol. C, H, N analyzed values are C = 37.85% (37.24%), N = 9.13% (9.65%) and H = 4.01% (4.13%). The IR peaks with tentative assignments ( $\nu_{\text{max}}/\text{cm}^{-1}$ ) at 1647(C=N aromatic), 3116(C-H,  $\text{SP}^2$  Carbon), 2920, 2822 (C-H methyl), 1496, 1415(C=C for side chain and aromatic), 454 (Ru-N) were observed.

### Crystal structure

Fine crystals were mounted on glass capillary and data collected at room temperature. Data collection was carried out on a diffractometer (BRUKER SMART APEX2) using monochromated Mo  $\text{K}\alpha$  radiation. From the setting angles crystal dimension were determined. The structures (Figure 1) were solved by the Patterson method. SHELXTL routine was used for empirical absorption correction. Experimental details are given in Table 1.



**Figure 1.** ORTEP diagram of  $[\text{Ru}(\text{Blm})_2\text{D}_2\text{Cl}_2] \cdot \text{H}_2\text{O}$ . (Blm=Benzimidazole, D=DMSO)

**Table 1.** Crystal data and structure refinement for *cis*-dichlorobis-(benzimidazole)bis-dimethylsulfoxideruthenium(II) hydrate, [Ru(BIm)<sub>2</sub>D<sub>2</sub>Cl<sub>2</sub>].H<sub>2</sub>O complexes

Identification code	[Ru(BIm) <sub>2</sub> D <sub>2</sub> Cl <sub>2</sub> ].H <sub>2</sub> O	
Empirical formula	C <sub>21</sub> H <sub>28</sub> Cl <sub>2</sub> N <sub>4</sub> O <sub>3</sub> Ru S <sub>2</sub>	
Formula weight	620.56	
Temperature	296(2) K	
Wavelength	0.71073 Å	
Crystal system, space group	triclinic, P-1	
Unit cell dimensions	a = 9.6901(10) Å	α = 73.701(4)deg.
	b = 11.1126(12) Å	β = 80.993(4)deg.
	c = 11.7112(12) Å	γ = 80.011(4)deg.
Volume	1184.3(2) Å <sup>3</sup>	
Z, Calculated density	2, 1.740 Mg/m <sup>3</sup>	
Absorption coefficient	1.097 mm <sup>-1</sup>	
F(000)	632	
Crystal size	0.07mm x 0.13mm x 0.24 mm	
Theta range for data collection	1.93 to 27.66 deg.	
Limiting indices	-12 ≤ h ≤ 12, -14 ≤ k ≤ 14, -15 ≤ l ≤ 14	
Reflections collected / unique	23879 / 5471 [R(int) = 0.0393]	
Completeness to theta	= 27.66 99.0 %	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	5471 / 0 / 278	
Goodness-of-fit on F <sup>2</sup>	1.119	
Final R indices [I > 2σ(I)]	R <sub>1</sub> = 0.0367, wR <sub>2</sub> = 0.1152	
R indices (all data)	R <sub>1</sub> = 0.0384, wR <sub>2</sub> = 0.1173	
Extinction coefficient	0.0051(11)	
Largest diff. peak and hole	1.020 and -1.698 e.Å <sup>-3</sup>	

## Results and Discussion

### *Spectroscopic studies on DNA binding: UV-Visible absorption titration*

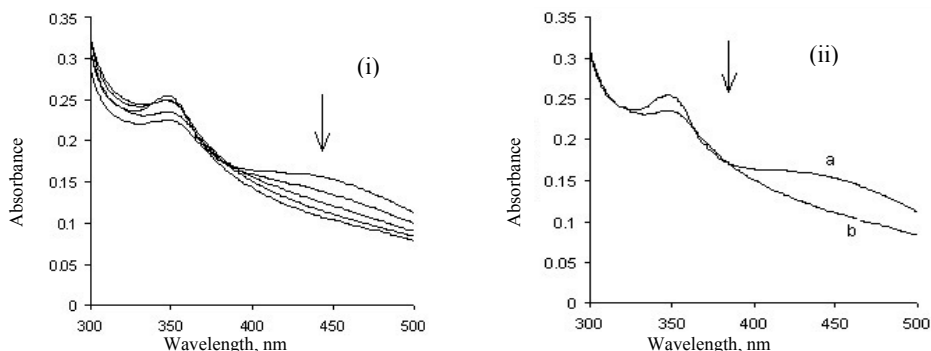
Interaction of the complexes with plasmid-DNA was monitored by UV-Visible absorption spectra of the complex at different concentration of DNA (2.289 × 10<sup>-6</sup> M to 9.089 × 10<sup>-6</sup> M). As the concentrations of the plasmid-DNA increased, red shift occurred from 351 nm to 359 nm and the absorbance decreases (Figure 2). The binding strength was estimated from the intrinsic binding constant, from the ratio of slope and intercept of the graph (Figure 3) from the following equation (1),

$$[\text{DNA}] / (\epsilon_a - \epsilon_f) = [\text{DNA}] / (\epsilon_b - \epsilon_f) + 1/(K (\epsilon_b - \epsilon_f)) \quad (1)$$

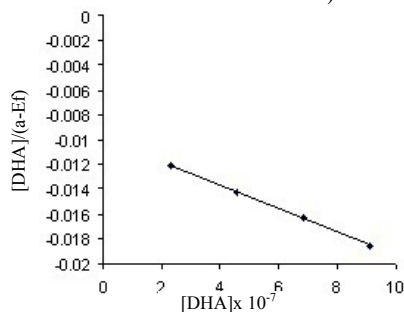
where  $\epsilon_a$ ,  $\epsilon_f$  and  $\epsilon_b$  are the extinction coefficient of observed solution, free complex and the complex when it fully bound to plasmid-DNA respectively.

### *Fluorescence emission and quenching studies*

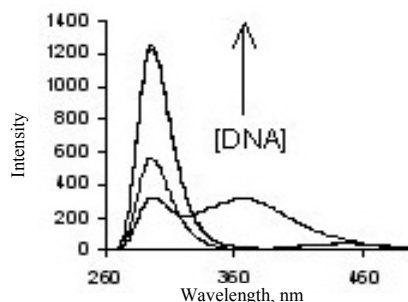
Fluorescence study was performed in two phases, firstly keeping the concentration of metal complex constant and varying concentration of CT-DNA (Figure 4) and secondly concentration of CT-DNA is fixed, and varying the concentration of metal complex (Figure 5). Steady-state emission quenching experiments were performed using [Fe(CN)<sub>6</sub>]<sup>4-</sup> as quencher and the Stern–Volmer quenching constant (K<sub>sv</sub>) was calculated by using Stern–Volmer equation<sup>12</sup>  $I_0 / I = 1 + K_{sv} [Q]$  (Figure 6).



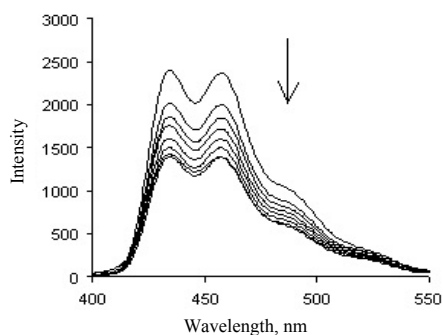
**Figure 2.** (i) UV-Visible spectra of  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  in tris-buffer (pH=7.4) with increasing concentrations of plasmid-DNA and (ii) with maximum concentration of plasmid-DNA indicating red shift from 351 nm (a) for the complex without DNA to 359 nm (b) with maximum concentration of DNA)



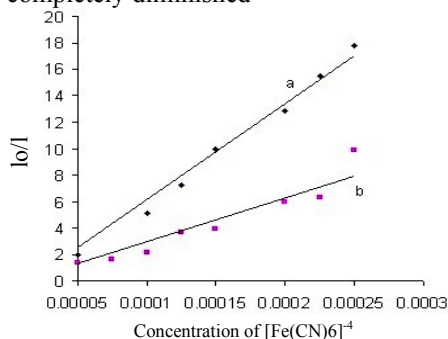
**Figure 3.** Plot of  $[\text{DNA}]/(a-ef)$  vs.  $[\text{DNA}]$ . Binding constant =  $9.09 \times 10^5 \text{ M}^{-1}$



**Figure 4.** Fluorescence spectra of  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$ ,  $10^{-3} \text{ M}$  with different concentration of DNA (0 to  $9.089 \mu\text{M}$ ). Exciting wavelength = 220 nm. Intensity at 300nm increases but at 370 nm intensity completely diminished



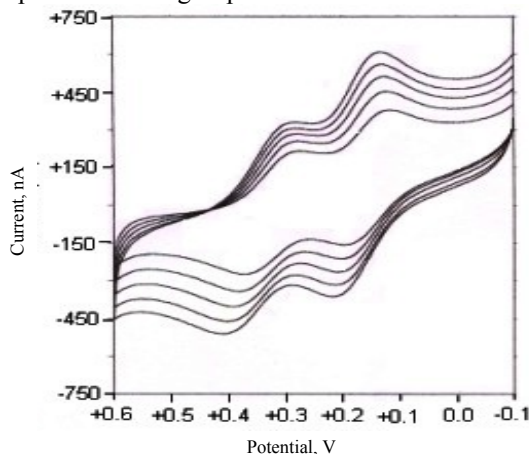
**Figure 5.** Fluorescence spectra of plasmid-DNA with different concentration of  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  (0 to  $7.6 \times 10^{-4} \text{ M}$ ). Exciting wavelength = 220 nm. Intensity decreases with increasing concentration of drug



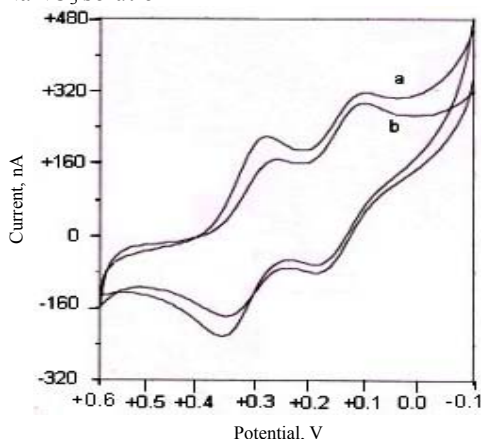
**Figure 6.** Stern-Volmer plot a) without plasmid DNA and b) with 30  $\mu\text{L}$  of 0.1 mM plasmid DNA. Slope of line a is 72079 and that of line b is 33131

### Electrochemistry

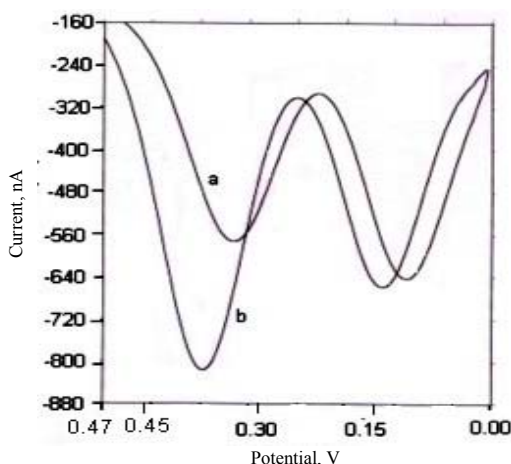
Cyclic voltammetric study for  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  complexes were carried out in water/ethanol solution containing 0.1M  $\text{NaNO}_3$  as supporting electrolyte using  $\text{Ag}/\text{Ag}^+$  as reference electrode, and a glassy carbon was used as working electrode<sup>1</sup>. The inert environment was maintained by passing  $\text{N}_2$  gas through the solution to remove oxygen. The voltamogram shows distinct oxidation and reduction peaks of a reversible electron transfer reaction. The shifts of redox potentials due to binding with CT-DNA are shown in Figure 7, 8 and 9. Interacting with DNA, by intercalative mode increases the hydrophobicity of the complex. As a result there is a positive shifting of potential.



**Figure 7.** Cyclic voltammogram of  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  at different scan rate (50 mV/s to 350 mV/s) in water/0.1M  $\text{NaNO}_3$  solution



**Figure 8.** Cyclic voltammogram of  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  (a) without Plasmid-DNA (Segment I :  $E_1 = +174$  mV, current =  $-8.296 \times 10^{-8}$  A and  $E_2 = +361$  mV; current =  $-1.069 \times 10^{-7}$  A; Segment II:  $E_1 = +103$  mV, current =  $+1.324 \times 10^{-7}$  A and  $E_2 = +268$  mV, current =  $+1.626 \times 10^{-7}$  A) and (b) with 60  $\mu\text{L}$  0.5 mM plasmid-DNA. (Segment I:  $E_1 = +169$  mV, current =  $-6.366 \times 10^{-8}$  A and  $E_2 = +369$  mV; current =  $-1.696 \times 10^{-7}$  A; Segment II:  $E_1 = +105$  mV, current =  $+1.270 \times 10^{-7}$  A and  $E_2 = +292$  mV; current =  $+1.984 \times 10^{-7}$  A)



**Figure 9.** OSWV of (a)  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$ ,  $(E_{1/2})_1 = +135$  mV; current =  $-6.505 \times 10^{-7}$  A and  $(E_{1/2})_2 = 321$  mV, current =  $-5.121 \times 10^{-7}$  A; (b)  $[\text{Ru}(\text{BIm})_2\text{D}_2\text{Cl}_2]$  with 30  $\mu\text{L}$  0.5 mM plasmid-DNA.  $(E_{1/2})_1 = 142$  mV; current =  $-3.905 \times 10^{-7}$  A and  $(E_{1/2})_2 = 331$  mV; current =  $-5.697 \times 10^{-7}$  A. Scan rate = 50 mV/s.

## Conclusion

The structural analysis was performed by single crystal XRD study, which shows six coordinated octahedral geometry. The results characterized by various techniques give evidences binding of the complex with plasmid-DNA. On the basis of spectroscopic shift, the intrinsic binding constant ( $K_b$ ) is  $9.09 \times 10^5 \text{ M}^{-1}$ . Emission quenching with  $[\text{Fe}(\text{CN})_6]^{4-}$  in presence and absence of plasmid-DNA shows the Stern-volmer constant 33131 and 72079 respectively. Based on the observed distinct electrochemical  $E_{1/2}$  and electrophoresis band shifts, the complex is expected to bind with plasmid-DNA.

## References

1. Mylonas S, Valavanidis Y A, Dimitropoulos K, Polissiou M, Tsiftoglou A S and Vizirianakis I S, *J Inorg Biochem.*, 1988, **34(4)**, 265-275; DOI:10.1016/0162-0134(88)83004-6
2. Haga M, Ali M M and Arakawa R, *Angew Chem., Int Ed Engl.*, 1996, **35(1)**, 76-78; DOI:10.1002/anie.199600761
3. Nial J Wheate, Shonagh Walker, Gemma E Craig and Rabbab Oun, *Dalton Trans.*, 2010, **39**, 8113-8127; DOI:10.1039/C0DT00292E
4. Gava B, Zorzet S, Spessotto P, Cocchietto M and Sava G, *J Pharmacology Experimental Therapeutics*. 2006, **317(1)**, 284-291.
5. Naishadham P, Pallavi P, Reddy K L and Satyanarayana S, *J Chem Pharm Res.*, 2012, **4(6)**, 3309-3318.
6. Rillema D P, Sahai R, Matthews P, Edwards A K, Shaver R J and Morgan L, *Inorg Chem.*, 1990, **29(2)**, 167-175; DOI:10.1021/ic00327a006
7. Haga M, Ali M Md, Koseki S, Fujimoto K, Yoshimura A, Nozaki K, Ohno T, Nakajima K and Stufkens D J, *Inorg Chem.*, 1996, **35(11)**, 3335-3347; DOI:10.1021/ic950083y

8. Haga M, Ali M Md, Maegawa H, Nozaki K, Yoshimura A and Ohno T, *Coord Chem Rev.*, 1994, **132**, 99-104; DOI:10.1016/0010-8545(94)80028-6
9. Haga M, Ali M M and Arakawa R, *Angew Chem., Int Ed Engl.*, 1996, **35**(1), 76-78; DOI:10.1002/anie.199600761
10. Kabanos T A, Kersmidas A D, Mentzafos D, Russo U, Terzis A and Tsangaris J M, *J Chem Soc., Dalton Trans.*, 1992, **19**, 2729-2733.
11. Biddle B N and Gray J S, *Appl Organomet Chem.*, 1991, **5**(5), 439-441; DOI:10.1002/aoc.590050512
12. Hazarika P, Deka J, Bhola S, Bhola R K, Medhi C and Medhi O K, *Int J Drug Des Dis.*, 2012, **3**(4), 907-913.
13. Nial J Wheate, Shonagh Walker, Gemma E Craig and Rabbab Oun, *Dalton Trans.*, 2010, **39**, 8113-8127; DOI:10.1039/C0DT00292E
14. Gorakh S Yellol, Antonio Donaire, Jyoti G Yellol, Vera Vasylyeva, Christoph Janiakb and Jose' Ruiz, *Chem Commun.*, 2013, **49**, 11533-11535; DOI:10.1039/C3CC46239K
15. Kalyanasundaram K and Nazeeruddin Md K, *Inorg Chim Acta*, 1994, **226**(1-2), 213-230; DOI:10.1016/0020-1693(94)04089-3
16. Barton J K.; Dannenberg J J and Raphael A L, *J Am Chem Soc.*, 1982, **104**(18), 4967-4969; DOI:10.1021/ja00382a048 (b) Barton J K, *J Biomol Struct Dyn.*, 1983, **1**(3), 621-632; DOI:10.1080/07391102.1983.10507469
17. Ward M D, *Chem Soc Rev.*, 1995, 121-134; DOI:10.1039/CS9952400121
18. Vajpayee V, Lee S M, Park J W, Dubey A, Kim H, Timothy R Cook, Stang P J and Chi K W, *Organometallics*, 2013, **32**(6), 1563-1566; DOI:10.1021/om301174s
19. Chaires J B, Dattagupta N and Crothers D M, *Biochemistry*, 1982, **21**(17), 3927-3932; DOI:10.1021/bi00260a004
20. Joseph R and Lakowicz G W, *Biochemistry*, 1973, **12**(21), 4161-4170; DOI:10.1021/bi00745a020
21. Veyisoğlu F, Mutlu A, Şahin, Nuran B, Pekmez O, Can A M, Attila C A and Yıldız, *Acta Chim Slov.*, 2004, **51**, 483-494.
22. Barton J K, Danishefsky A T and Goldberg J M, *J Am Chem Soc.*, 1984, **106**(7), 2172-2176; DOI:10.1021/ja00319a043
23. Lakowicz J R Principles of fluorescence spectroscopy, Plenum Press York, 1983.