

Synthesis, Characterization and Antimicrobial Screening of Fe(II)-Glycine Schiff Base

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Abstract: Fe(II) complex of (*E*)-2-((2-hydroxynaphthalene-1-yl)methyleneamino)acetic acid was synthesized by reacting Fe(OAc)₂ with Schiff base ligand in stoichiometric 1:1 ratio and was characterized by elemental analysis, IR, Mass and conductance measurement. Furthermore, this complex was screened for antimicrobial activity against *E. coli*, *S. aureus*, *A. niger* and *C. albicans*.

Keywords: 2-Hydroxy-1-naphthaldehyde, Glycine, Schiff Base, Fe(II) complex, Antimicrobial activity

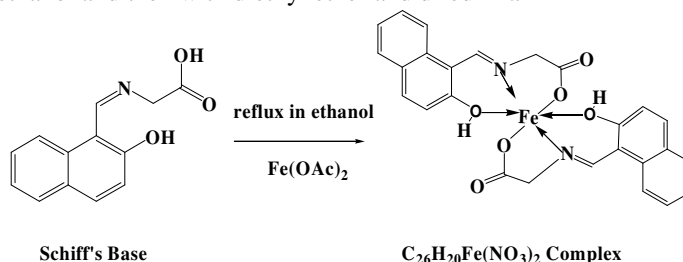
Introduction

In 1864, German chemist Hugo Schiff developed a new class of organic compounds¹. The active and well-designed Schiff base ligands are considered as "Privileged Ligands" by Cozzi². Schiff base ligands were synthesized by condensation of α -amino acid with aldehyde to form azomethine ($-\text{RC}=\text{N}-$) linkage. The metal complexes with Schiff bases play important role in biology^{3,4}, analytical chemistry^{5,6} and industry^{7,8}. The complexes of amino acid Schiff bases prepared from *o*-hydroxy aryl aldehydes were used as radiotracers in nuclear medicine, antibacterial and anticancer agents^{4,9-11}. Bioinorganic chemistry is important and has great impact in coordination chemistry¹². Haemoglobin is Fe(II) containing complex acts as a oxygen carrier. Ca(II) complexes is basic constituents of bone, Zn²⁺ ions found in three-dimensional structural framework of proteins are good examples of metal complexes in biological systems¹³. Metal ions act as redox transfer and hence metal complexes showed medicinal applications like organic compounds which could be used in drug discovery efforts¹⁴. They were used as therapeutic agents since 3500 BC¹⁵. Over the past

Scheme 1. Synthesis of 2-hydroxy-1-napthaldehyde based Schiff base

Synthesis of complex $C_{26}H_{20}Fe(NO_3)_2$

Iron acetate (0.01 mol) in anhydrous ethanol (15 mL) was added in stirring solution of Schiff base (0.01 mol) in 15 mL anhydrous ethanol. The resulting solution was stirred overnight at 70 °C (Scheme 2). The dark brown precipitate obtained was filtered, washed with ethanol and then with diethyl ether and dried in air



Scheme 2. Complex Formation Reaction of Schiff's base with transition metal

Spectral data for complex ($C_{26}H_{20}Fe(NO_3)_2$)

IR(KBr) ν cm^{-1} : 1610(C=N), 1367(COO⁻), 561(M-N), 488(M-O) MS(m/z): 512 *Anal.* Calcd. for C-60.96, H-3.94, N- 5.47, Fe-10.90, found C- 63.60, N-4.30, Fe- 10.77

Materials and Methods

All reagents and solvents used were of analytical grade and were used without further purification. 2-Hydroxy-1-naphthaldehyde, glycine, iron acetate was purchased from Sigma Aldrich, Merck and Spectrochem chemicals. Melting points were determined on a Gallenkamp melting point apparatus (Table 1). The ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Avance II 500 MHz Spectrometer. Chemical shifts were reported in ppm relative to tetramethylsilane (TMS), and multiplicities are given as s(singlet), bs(broad singlet), d(doublet), t(triplet), q(quartet), or m(multiplet). Infrared spectra were recorded as KBr pellets on a Shimadzu FTIR-408 spectrophotometer. EDS analysis was performed at CIF, SPPU, Pune-7. Mass spectra were recorded on a Shimadzu LC-MS:EI QP 2010A mass spectrometer with an ionization potential of 70 eV. Molar conductivity of complexes was recorded using 1×10^{-3} M solutions in DMSO on Toshniwal TSM 15 conductivity meter. Reactions were monitored by thin layer chromatography (TLC), carried out on 0.2 mm silica gel 60 F₂₅₄ Merck plates using UV light (254 and 366 nm) for detection.

Results and Discussion

Molar conductance measurements

The value of molar conductance of complex $C_{26}H_{20}Fe(NO_3)_2$ when dissolved in DMSO was observed $15.65 \Omega^{-1}cm^2mol^{-1}$ respectively indicated the non-electrolytic nature of this complex.

Table 1. Reaction time, % yield, color and melting point of ligand and complex

Mol. formula	Ligand/Complex	Reaction time, h	Yield, %	Colour	M.P. °C
C ₁₃ H ₁₁ NO ₃	Ligand	4	88	Yellow	240
C ₂₆ H ₂₀ Fe(NO ₃) ₂	Complex	12	80	Brown	242

IR Spectra

IR Spectra of ligand (Figure 1) showed band at 1641 cm^{-1} which is due to ($\nu C=N$) confirming the formation of Schiff base. The band of ($\nu C=N$) at 1641 cm^{-1} in ligand was shifted to 1610 cm^{-1} in complex which indicate the coordination of azomethine group through its nitrogen atom²⁵.

The band at 1604 and 1394 cm^{-1} in spectrum showed the presence of COO^- , these bands were shifted to 1427, 1367 cm^{-1} in Fe(II) complex indicated metal-oxygen coordination. In complex the difference $[\Delta\nu(\text{COO})]$ between $\nu_{\text{assy}}(\text{COO})$ and $\nu_{\text{sy}}(\text{COO}) > 200 \text{ cm}^{-1}$ indicated the unidentate coordination of COO^- with Fe(II) metal²⁶. The band at 561 cm^{-1} indicated the coordination of azomethine through nitrogen with metal ion, while band at 488 cm^{-1} was due to (M-O) stretching in the complex²⁷. The IR band at 3414-3340 cm^{-1} in complex corresponded to phenolic $-\text{OH}$ stretching. This stretching frequency shift indicates the coordinate bonding of $-\text{OH}$ oxygen to the metal²⁸. Thus the ligand coordinated to free metal ion through its azomethine($-\text{RC}=\text{N}-$) nitrogen and oxygen of (COO^-) carboxylate and O of the phenolic $-\text{OH}$ indicated tridentate nature of ligand.

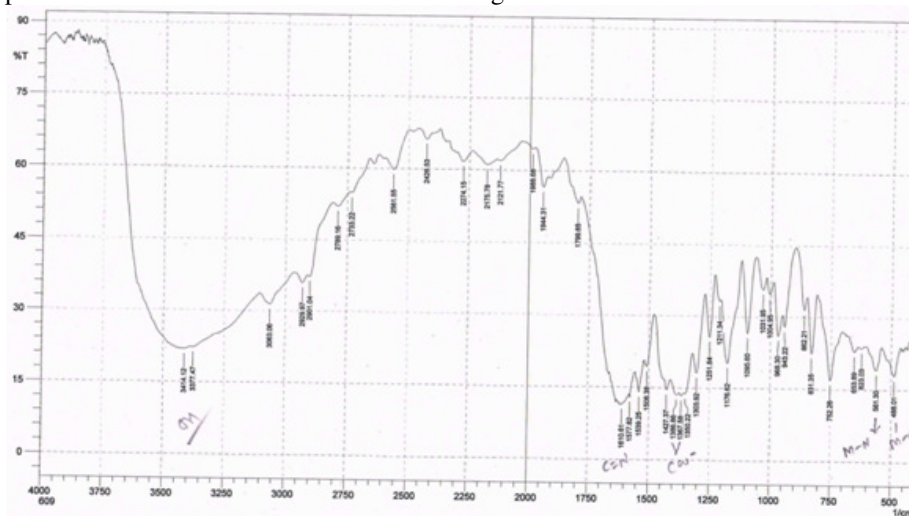


Figure 1. IR Spectra of complex, $\text{C}_{26}\text{H}_{20}\text{Fe}(\text{NO}_3)_2$

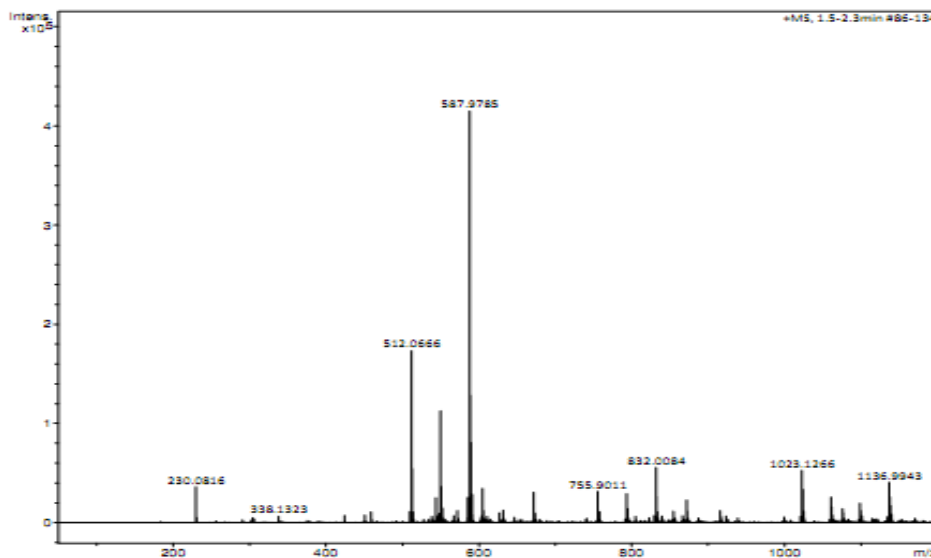


Figure 2. Mass spectrum of complex, $\text{C}_{26}\text{H}_{20}\text{Fe}(\text{NO}_3)_2$

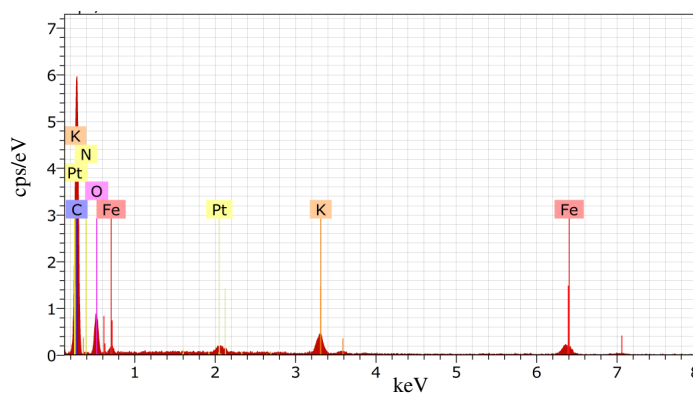


Figure 3. EDS of complex, $C_{26}H_{20}Fe(NO_3)_2$

Mass spectrum of complex

Mass spectra of complex (Figure 2) shows M^+ peak at 512.06 corresponded to molecular formula $C_{26}H_{20}Fe(NO_3)_2$.

EDS of complex

EDS of complex shows 10.77% of Fe(II), which is in good agreement with elemental analysis (Figure 3).

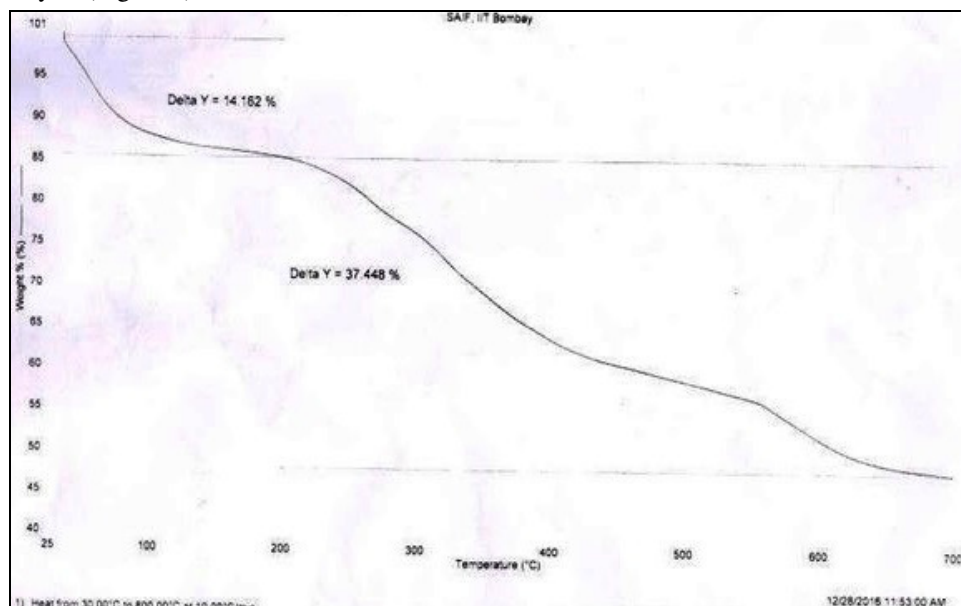


Figure 4. TGA of complex $C_{26}H_{20}Fe(NO_3)_2$

TGA

The TGA curve of synthesized Fe complex (Figure 4) reveals weight loss occurring in two stages. The first small weight loss is due to absorbed moisture which is found at 100 °C and the second exothermic loss of 37.448% due to decomposition of Fe complex and crystallization of FeO particles. The theoretical weight loss calculated by conversion of

complex precursor to FeO is in good agreement with the measured value. The process accounts for one third of total weight loss in the whole decomposition process from 100 to 600 °C according to TGA results, with the major loss occurring between 200 to 500 °C. The TGA result shows that complex is highly stable up to 225 °C.

Antimicrobial assay

The antimicrobial assay of Fe metal complex was done using agar well plate method (Figure 5). The antibacterial and antifungal assays were performed in Muller-Hinton broth and Crazeck Dox broth³⁰. The standard strains used was procured from Microbial Culture Collection, Pune, India. Antimicrobial evaluation was performed using the bacteria reseeded in Muller-Hinton broth for 24 h at 37 °C and fungi reseeded in Crazeck Dox broth³⁰ for 48 h at 25 °C. The antibacterial activity of tested samples were studied (Table 2) in triplicate against gram positive bacteria *Staphylococcus aureus* (ATCC 29737) and gram negative bacteria *Escherichia coli* (ATCC 25922). The same samples were tested for antifungal activity in triplicate against *Candida albicans* (MTCC 277) and *Aspergillus niger* (MCIM 545). The compounds were dissolved in DMSO at desired concentrations of 40, 20, 10 µg/mL. DMSO was loaded as negative control. Gentamicin (10 µg/mL) and Fluconazole (20 µg/mL) were used as standards for evaluating the antibacterial and antifungal activity. The zone of inhibition (mm) was determined from the diameter of the zone of inhibition using caliper as per National Committee for Chemical Laboratory Standards (NCCLS, M7-A5, January 2000). Both metal complexes shows good antibacterial as well as good antifungal activity.

The complex $C_{26}H_{20}Fe(NO_3)_2$ showed excellent antibacterial activity against *Escherichia coli* (ATCC25922) with MIC 20 µg/mL when compared with standard antibacterial drug gentamicin (10 µg/mL). Similarly it also showed excellent antifungal activities against *Aspergillus niger* (MCIM 545), *Candida albicans* (MTCC 277) with MIC 10 µg/mL when compared with standard antifungal drug fluconazole (20 µg/mL) (Table 3).

Table 2. Antimicrobial screening of complex $C_{26}H_{20}Fe(NO_3)_2$

Complex	Inhibition Zone Diameter, mm			
	<i>E. coli</i> (ATCC25922)	<i>S. aureus</i> (ATCC 29737)	<i>A.niger</i> (MCIM 545)	<i>C.albicans</i> (MTCC 277)
$C_{26}H_{20}Fe(NO_3)_2$	15±0.3	22±0.5	21±0.8	19±0.8
DMSO	11±0.7	12±0.9	12±0.6	13±0.3
Gentamicin	22±0.4	23±0.7	-	-
Fluconazole	-	-	23±0.8	24±0.5

Gentamicin (10 µg/mL) and fluconazole (20 µg/mL) Inhibition Zone= 9-14 mm: slight activity, 15-19 mm: moderate activity, 20 -24 mm : high activity, >25 mm: excellent activity.

Table 3. Antimicrobial screening of complex $C_{26}H_{20}Fe(NO_3)_2$

Complex	Minimum Inhibition Concentration (MIC), µg/mL			
	<i>E.coli</i> (ATCC25922)	<i>S.aureus</i> (ATCC 29737)	<i>A.Niger</i> (MCIM 545)	<i>C.albicans</i> (MTCC 277)
$C_{26}H_{20}Fe(NO_3)_2$	10	10	10	10
Gentamicin	10	10	-	-
Fluconazole	-	-	20	20

Gentamicin (10 µg/mL) and Fluconazole (20 µg/mL), (MIC in µg/mL)=10 µg/mL: excellent activity, 20 µg/mL: moderate activity, 40 µg/mL: slight activity

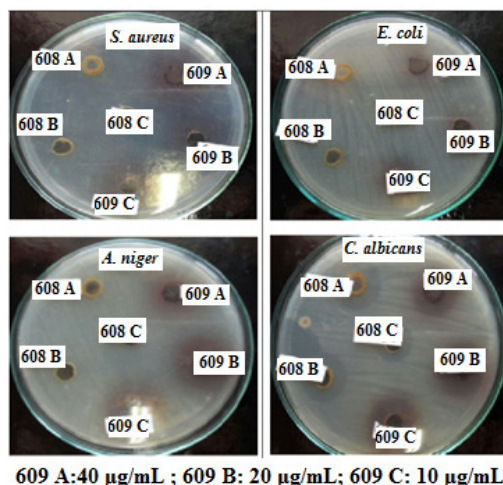


Figure 5. Antimicrobial screening of complex (609): $C_{26}H_{20}Fe(NO_3)_2$

Conclusion

The synthesis and characterization of Fe(II) Complex were studied herein in the light of elemental analysis, 1H NMR, IR, electronic spectra and mass spectra. It was concluded that the Schiff base coordinating through the azomethine nitrogen and carboxylate oxygen. The synthesized complex has good antimicrobial activities.

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